

Sugden Lab.

SI Nuclease Mapping Protocol

Kinase of Oligonucleotides

γ 32P ATP ~6,000Ci/mM, 150 mCi/ml CAT#NEG-035C (DuPont NEN)
1ul Oligo @ 10pm/ul 32P ATP comes at ~20pm/ul
2ul 10xNEB Kinase Buffer Use at 2:1 ratio of [ATP] / [Oligo]
15ul H2O
1ul 32P ATP
1ul PNK

On ice o/n @ 4°C

Qiagen Nucleotide Removal Kit (50% recovery in 50ul) or a G25 column
Oligonucleotide will be at 100 fm/ul final concentration

Annealing and SI Digestion

50 µg of RNA for Luciferase Assays
5 or 25 µg of RNA for GAPDH
100 fm of each oligonucleotide

	Final	SI Buffer (2mL)
20 µl Formamide	50%	1 µl SI Nuclease (400Units)
3.2 µl 5M NaCl	0.4M	100 µl 5M NaCl
1.6 µl 1M PIPES (pH 6.4)	0.04M	20 µl 3M NaOAc
4 µl 1% SDS	0.1%	20 µl 100mM ZnSO ₄
1 µl labeled Oligonucleotide DNA	2.5fm/µl	1µl 10 mg/ml Salmon Sperm
x µl RNA		1860µl H ₂ O
up to 40 µl with H ₂ O		

Anneal at 42°C >12 hours

400 µl SI Buffer Mix

50°C 75-90min

Stop reaction by adding 80µl 4M NH₃OAc w/20mM EDTA and 3.2µg of tRNA
(400 µl 4M NH₃OAc + 16 µl 0.5M EDTA + 1 µl 15mg/ml tRNA+4 µl Glycogen)

EtOH PPT with 1ml 100% EtOH and resuspend in 5-10µl loading dye

Run on a 10% Dneanturing PAGE (23g UREA, 5ml 10xTBE, 12.5ml 40% 19:1
Acrylamide, 10 ml H₂O- dissolve and add 25 µl TEMED and 250 µl 10% APS)

Last Modified 06/05/05

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10x Poly Nuclotide Kinase Buffer (PNK)	1M PIPES (pH 6.4)	Loading Dye (1ml)
7 μ l 1M TRIS pH7.6	3g PIPES	950 μ l Formamide
1 μ l 1M MgCl ₂	9ml H ₂ O	40 μ l 0.5M EDTA
1 μ l 0.5M DTT	pH w/HCl	10 μ l of 5%B-Blue &
91 μ l H ₂ O	Use dipotassium of PIPES	Xylene Cyanol FF